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Advances in the Development of Antimicrobial Agents for Textiles: The Quest for Natural Products. Review

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Abstract

The antimicrobial finishing of textiles has attracted research attention lately due to demands for a healthy lifestyle. As a result, several synthetic and natural antimicrobial agents for textiles have been developed over the years. Recently research into antimicrobials agents of natural origin have become more popular due to their enormous therapeutic potential and effectiveness in the treatment of infectious diseases while mitigating the side effects of the synthetic antimicrobials. Research into these natural biocides for textiles has seen increasing consumers awareness for two reasons, namely the potential negative impact of synthetic biocides on health versus the benefits of natural biocides, and the increasing rate of microbial resistance to most natural biocides. The immense literature on natural biocides suggests the preparedness of the research community and industry in addressing the environmental and health challenges associated with synthetic antimicrobial agents in response to the new consumer demands. This review focuses on the advances in natural antimicrobial agents and various methods of their application. Literature suggest that natural antimicrobial agents have chalked some success in terms of efficacy and wash durability, with minimal effect on the tensile strength of fabrics.

Key words: antimicrobial finishing, synthetic antimicrobial agent, natural antimicrobial agent, textiles, essential oils, dyes.

Introduction

Fibres have been widely used in all areas of human activity as costumes, tents, tarpaulins, awnings, blinds, parasols, shower curtains, mattress ticking sails, waterproof clothing, etc. and in medical environments [1, 2]. However, natural fibres are prone to microorganism colonisation due to their large surface area and excellent moisture retention ability. Consumers are therefore justified in their increasing demand for the protection of fibres against microbes.

Antimicrobial finishing for textiles dates back to the ancient Egyptians, who applied herbs and spices to preserve mummy wrapping [3]. Today many different synthetic and natural compounds such as triclosan, quaternary ammonium, polybiguanides, N-halamines, silver and chitosan have been developed to impart antimicrobial properties to textiles [4 -7]. Estimates have shown that the production of antimicrobial textiles was in the magnitude of 100,000 tons worldwide in the year 2000 [4]. Antimicrobial textile production has soared over the years, making it one of the fastest growing sectors in the textile industry [8]. A market survey report released recently by Textile Intelligence forecasts the global market for antimicrobial agents for all end uses, including textiles, to grow by nearly 12% per annum between 2013 and 2018 [9]. This projected growth coupled with the increasing demand for safe antimicrobial products is bound to advance the cause of cleaner and environmentally friendly antimicrobial textiles.

Antimicrobial agents are usually applied at the finishing stages of textile production, while in some cases biocide can be incorporated into synthetic fibres during extrusion. Bacterial resistance to biocides, their toxic effects on households and the environment, inadequate activity and poor durability on textiles have, however, become important issues of concern [4, 10, 11]. As a result, certain synthetic antimicrobial agents such as triclosan have been banned by a number of leading retailers and governments in Europe, especially those with the potential to cause skin irritation, and nonbiodegradable and bioaccumulation effects [2]. Due to the high level of consumer awareness about clothing safety, many kinds of eco-friendly antimicrobial agents such as peroxy acids, chitosan and its derivatives, specific dyes and regenerable active N-halamine have been developed for textiles [12 - 15].

Lately investigation into natural antimicrobial agents, such as extracts from *neem*, *aloe vera*, *eucalyptus*, *capsaicin*, etc., have received great attention due to their eco-friendliness [10, 13, 16]. It has been found that natural antimicrobial

agents exhibit broad-spectrum activity against bacteria, fungi and viruses, but they still has some limitations in terms of efficacy and durability [17 - 19], hence the quest for suitable methods of application of these natural biocides to impart high efficiency and durability.

Requirement of antimicrobial agents for textiles

Antimicrobial agents improve resistance against microorganisms, increase fabric durability and protect textiles against the colonisation of odour forming bacteria [20, 21]. This addresses hygiene needs in clinical and sensitive environments by minimising the chances for the microbial colonisation of textiles [22]. Antimicrobial treatments can also reduce the frequency of laundering, which gives the potential for significant savings in water usage and energy consumption and reduces the need for chemical materials in textile care [10, 13]. A good antimicrobial agent should be effective against a broad spectrum of bacterial and fungal species and must also exhibit low toxicity to consumers and the environment while remaining allergy and irritation-free [11]. Textiles treated with an antimicrobial agent should not only meet the standards of cytotoxicity, irritation and sensitisation tests, but should also satisfy the requirement as suggested by [2, 23 - 25] as follows:

- a) Withstand repeated washing during its lifespan.
- b) Should not have a negative effect on the quality (e.g. physical strength and handle) or appearance of the textile.
- c) Be compatible with chemical materials for textile finishing.
- d) Should not kill the resident flora of non-pathogenic bacteria on the skin of the wearer.
- e) Comply with the statutory requirements of regulatory agencies.
- f) Should not produce side effects for the manufacturer, user and environment.
- g) Easily applicable and resistant to disinfection/sterilization.

Antimicrobial agents purported to be eco-friendly have been marketed commercially by some companies, for example, Ultrafresh (Thomsan Research Associates), 2, 4, 4'-Trichloro-2' - hydroxyldipenylether (Tinosan AM 110, Ciba Specialty Chemicals) [26], Sanitized AG (Clariant) [27], Ecosy (Unitika), Utex (Nantech Textile Company Limited) and Vantocil IB (Zeneca) [28] etc. However, a cursory look into the chemistry behind these purportedly natural biocides shows they are not entirely natural [29, 30]. Today wholly natural antimicrobial agents are rarely found on the market due to their poor efficacy and durability.

Natural antimicrobial agents

Research into natural products has demonstrated significant progress in the discovery of new compounds with antimicrobial activity from natural products. Among the known sources of natural compounds with valuable antimicrobial activity are medicinal plants, marine and terrestrial organisms, including fungi and bacteria [26, 31 - 36]. Nonetheless there is still vast fauna and flora that could provide antimicrobials for textiles. Several thousands of natural products with the potential to act as antimicrobial compounds still await further examination. Plants as a major source of natural antimicrobials in nature have received a lot of research attention lately [37 - 39]. Materials extracted from different parts of plants such as bark, leaves, roots and flowers containing common coloring materials such as tannin, flavonoids and quinonoids with strong antimicrobial properties, have been investigated [39, 40].

Extracts from plants

Several studies have shown that natural materials such as alkaloids, saponins, terpenoids and phenolic compounds possess strong antimicrobial activity [40 - 48]. Many of these natural antimicrobial agents are normally accumulated as secondary metabolites in plant cells, but their composition and concentration vary in different parts of plants [49]. Leaves have been found to contain the highest antimicrobial activity and are generally preferred for therapeutic purposes [50]. Perumal Samy et al., [51] found that vinyl hexylether and 2-methylnonane isolated from a Lonicera involucrata (Twinberry Honeysuckle) leaf had inhibitory activity against E. coli, and TIR-01(10,13dimethoxy-17-(6-methylheptan-2-yl)-2,-3-4,-7,-8,-9,-10,-11,-12,-13,-14,-15,-16,-17-tetradecahydro-1-Hcyclopenta TIR-05 (3-(2,4-dimethoxyphe-nyl)-6,7dimethoxy-2,3-dihydrochro-men-4-one) extracted from the twinberry honeysuckle root also possess good antibacterial property against E. coli [18, 51-58].

Gupta et al., [59] investigated the antimicrobial property of tannin-rich extract of the Quercus infectoria (QI) plant. They found that the use of copper sulphate and alum as a mordant together with the leaf extract had the potential to aid antimicrobial efficacy as well as durability. When the same extract was applied without the mordant, the antimicrobial activity was good but its durability was compromised because the antimicrobial activity decreased drastically after a few washes. They concluded that copper sulphate and alum could help improve the durability of antimicrobial activity of certain plant extracts in textiles.

Sathianarayanan et al., [60] extracted antimicrobial agents from Ocimum sanctum and the rind of Punica granatum, and compared the washing durability of fabrics treated with direct application, microencapsulation, resin cross-linking and their various combinations. Antimicrobial activities were evaluated using the AATCC-147 test method. Except direct application, all other treatments showed excellent antimicrobial activity as well as good wash durability. While a small decrease in the tensile strength and crease recovery angle was observed for resin treated and microencapsulated fabrics, no significant changes were observed in the combined processes [60].

Essential oils

Essential oils are a mixture of a variety of aromatic compounds which can give cologne and provide protection from a broad spectrum of microbes [61 - 63]. Due to their pleasant fragrance, essential oils containing different pharmaceutical effects have for centuries been applied on textiles to fulfill the psychological and emotional needs of humans [49, 64 - 66]. The application of essential oils for antimicrobial effect on textiles have increased in recent times because they are highly efficient when applied using the right technology [62, 65].

Although significant research has shown the mode of action of essential oils and their combinations for preservatives or antibiotics [67 - 79], the exact mechanism/mode of action of their synergies or purified components on microorganisms when applied in textiles have not been explicitly described [80 - 83]. Presently the generally accepted mechanism of antimicrobial action is the sequential inhibition of the common biochemical pathway, protective enzymes and the destruction of the cell wall of the microbe enhancing quick uptake of the antimicrobial agent as well as cell lysis [84 - 88]. For example, the synergistic effect of carvacrol and some hydrocarbon monoterpenes (such as α-pinene, camphene, myrcene, α-terpinene and p-cymene) showed good antimicrobial properties because the hydrocarbons interacted with the cell membrane of the microbes and facilitated quick penetration of carvacrol into their cells [89 - 91]. In a similar work carried out by Pei et al., [92] and confirmed by other researchers on the synergic effects of eugenol/carvacrol and eugenol/thymol, they intimated that carvacrol and thymol disintegrated the outer membrane of E. coli, making it easier for eugenol to enter the cytoplasm [94, 95]. The advantages of synergy with various essential oils is that it reduces the concentration needed to yield the same antimicrobial effect when compared with the sum of the purified components [93 - 96].

However, the synergistic effects of essential oils for textile applications, to the best of our knowledge, have not been reported, even though essentials oils have been extensively applied.

Matan et al., [97] showed that thyme essential oils have the ability to inhibit bacterial growth when applied using

the pad-dry-cure approach. Walentowska et al., [66] also applied thyme essential oil to increase the resistance of lignocellulose textiles to bacteria and mould action. However, other application techniques such as microencapsulation, nanotechnology and cross-linking have been reported [98, 99]. Peppermint oil, with its main component being menthol, is another essential oil with strong antimicrobial activity. It has been reported to exhibit high fungistatic and fungicidal activity against mould growth on rubber wood and other substrates [100 - 103]. Moreover essential oils from clove, cinnamon and several other oils have also been used as antimicrobial agents for bio-medical applications and in the food industry [50, 75, 104-106]. Excessive application of essential oils in linen-cotton blends have been found to affect the tensile strength of fabrics [107], hence the need for compromise between the biocidal efficiency and tensile strength.

Natural pigments

Natural dyes have been widely used in textile coloration since ancient times [19, 108 - 111]. One of the most significant benefits of natural dyes is the eco-friendliness, i.e., they do not create any environmental problems at the stage of production and maintain stable ecological balance [112]. Furthermore natural dyes have an inherent antimicrobial property which is believed to be very potent [110, 113-115]. Natural dyes extracted from different parts of plants including bark, leaves, roots, fruits or seeds, and flowers contain different colouring materials such as tannin, flavonoids and quinonoids [115, 116]. Some natural dyes come from microorganisms such as fungi, algae and bacteria. These dyes do not only offer rich and a wide range of colors, but also possess antimicrobial properties, are environmentally friendly and can be used in low-cost treatments with the additional benefit of colouring in a single step [40].

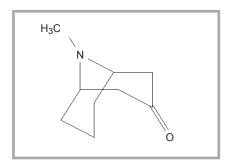


Figure 1. Chemical structure of granatonine.

As a result, research into the use of non-toxic, antimicrobial and eco-friendly natural dyes for textiles, preferably natural fibre products, has increased in recent times [117 - 119].

Calis et al., [120] showed the antimicrobial activity of natural dyes extracted from Rubia tinctorum, Allium cepa, Punica granatum and Mentha sp. They found that dye extract of Punica granatum was effective against all the bacteria tested except Escherichia coli and staphylococcus epidermidis. The textile material impregnated with the natural dyes above recorded a high rate of inhibition against bacteria [20]. Mirjalili et al., [121] demonstrated a one-step dyeing process using walnut shells to impart colour and a antimicrobial property to polyamides. Natural dyed polyamide fabrics exhibited good wash fastness and excellent antibacterial activity against two well-known pathogenic bacteria S. aureus and E. coli when the fabric was treated with a mordant [121]. Jafari et al., [122] also showed that the dyeing of soybean protein fibres with madder, weld and walnut seed husk could exhibit excellent antimicrobial effects against Gram-positive bacteria.

The use of fruits for dyeing and antimicrobial finishing is not a new phenomenon, however, rarely do we find literature directly linking the antimicrobial effects of fruits on microbes especially in textiles, with the exception of *pomegranate* rind, which have been reported recently [123 - 127]. The rind of *pomegranate* is a rich source of tannins, phenolic content, alkaloids, etc. *Granatonine* is the main colouring component found in fruit rind in the form of N- methyl *granatonine* (*Figure 1*).

The extract of pomegranate rind has been used in the dyeing and bactericidal treatment of cotton fabric [128, 129]. It offered a wide range of shades with significant antimicrobial activity when copper and iron were used as mordant [123, 128]. When pomegranate rind was used without the mordant (copper, iron). few shades of colours were developed. Moreover the antimicrobial activity of the dyed cotton was satisfactory but not durable enough to withstand repeated washing. The mordant (copper and iron) helped to fix the dye into the fabric, while improving the antimicrobial activity of the fabric because they already possess

some level of antimicrobial properties [128, 129].

The antimicrobial properties of pigment from microorganisms have also been investigated [130, 131]. Pigment from the fungi *Monascus purpureus* was extracted using ethanol and applied for dyeing wool fibres [131]. Excellent colour fastness with good antimicrobial properties was obtained when alum and stannic chloride were used as mordents. Kim et al., [132] also used *protease* extract from bacteria (*Rhizopus oryzae*) for dyeing wool and silk, and the result showed that by increasing the *protease* extract, the colour of the fabric was enhanced with a good antimicrobial property.

The antibacterial properties of pigments obtained from other fungal species, namely isaria farinosa, emericella nidulans, fusarium verticillioides and penicillium purpurogenum were studied [133 - 135]. The results showed that the dyed fibres had antimicrobial activity with excellent colour fastness [136]. The extraction, characterisation, and use of pigment produced by the bacteria Serratia sakuensis has also been investigated. A novel red pigment was produced by solid-state cultivation of bacterial isolate obtained from garden soil, and used for dyeing silk, wool and cotton fabrics. The dyed fabrics demonstrated good colourfastness and antibacterial activity [137]. Similarly Sharma et al., [138] applied pigments obtained from the fungi Trichoderma virens, Alternaria alternate and Curvularia lunata to wool and silk fibres without a mordant. The fibres showed good rubbing fastness with promising antifungal activity.

Chitosan and its derivatives

Chitosan and its derivatives appear to be the most effective natural antimicrobial agent on the market. Deacetylated chitosan derives its antimicrobial properties from the polycationic sites that are able to bind to negatively charged residues of macromolecules at the cell surface of the bacteria, which subsequently inhibit the growth of bacteria [13, 139 - 141].

Several mechanisms of action and modes of inhibition of chitosan have been suggested [142, 143], meanwhile there is no generally accepted mode of action because the dissimilar physical states and molecular weights of chitosan and its derivatives render distinctive modes

of antibacterial action [144]. According to Khor et al., [145] low molecular weight water-soluble chitosan and ultrafine nanoparticles could penetrate the cell wall of bacteria and combine with its DNA, thereby inhibiting the synthesis of mRNA and DNA transcription, while high molecular weight water-soluble chitosan and solid chitosan including larger size nanoparticles interact with the cell surface and alter cell permeability [146]. Others have suggested a situation where the particles interact with the cell and form an impermeable layer around it, thus blocking the transport of essential solutes into the cell [144, 147 - 149]. Irrespective of the mode of action of chitosan, it is highly effective when applied at higher concentration and at a relatively lower of pH 6.5 [13].

Besides the biocidal properties of chitosan on textiles, it also has several other advantages for coloration because the amine group present readily reacts with dyes for successful dyeing/printing [13, 150, 151]. Chitosan is mostly applied by the traditional pad-dry-cure process using a chitosan/citric acid mixture mainly on cotton fabrics, even though other techniques have been used to impart antimicrobial property to fabrics. The use of binders with chitosan for imparting antimicrobial activity to fabrics has also been reported [152 - 154]. In the case of the latter, it can be applied to all manner of fabrics due to the presence of the binder.

The use of chitosan and its derivatives on fibres seems to be the more realistic prospect since this product does not provoke any immunological response [155].

Methods of application for natural antimicrobial agents

Natural antimicrobial agents can be applied to textile by different methods such as pad-dry-cure, coating, spray and foam techniques. It can also be applied directly by adding the antimicrobial agent into the spinning fibre solutions [25, 156 - 158]. Some of the well-acclaimed methods for application of natural products for impacting antimicrobial finishing to fabrics are as folows [159 - 162]:

- Direct application techniques.
- Nanotechnology.
- Insolubilisation of the active substances in/on the fibre.

- Treating the fibre with resin, condensates or cross-linking agents.
- Microencapsulation of antimicrobial agents with the fibre matrix.
- Fibre surface modification.

Direct application technique

Direct application is one of the oldest methods of imparting antimicrobial agents to fabrics. This method is simple but usually cannot achieve satisfactory durability due to the poor attractive force between the fibres and antimicrobial agent, for example, herbal extracts applied directly to textile by pad-dry-cure or [163, 164]. Pad-dry-cure is the conventional technique for applying finishing agents to most textile materials. It is a simple and easy technique where fabric is immersed for about 5 - 10 minutes in an aqueous treating solution. The fabric is padded through squeeze rolls to give a specified wet pick-up, reported as a percentage weight of fabric (% o.w.f.), after which it is dried and cured for a specified time at a specified temperature [165]. Unlike the pad-dry-cure method, the exhaust process involves loading fabric into a bath, originally known as a batch, and allowing it to reach equilibrium with a solution, suspension or dye. Exhaust dyeing enables molecules to move from the solution onto the fibres until it is completely exhausted, after which the fabric is rinsed to remove any excess solution/ dyestuff [166, 167]

These two methods are fundamental to any other method for imparting permanent finishes to fabrics regardless of the technology involved.

Nanotechnology

The application of nanotechnology is ideal for improving the antimicrobial activity and wash durability of textiles [168, 169] Hasabo et al., [99] prepared nanoscale particles from Azadirachta indica (neem) extract and used them to treat cotton fabrics. Such fabrics showed excellent antimicrobial activity against Staphylococcus aureus and Escherichia coli. They also demonstrated that the antimicrobial activity of neem nanoparticle treated fabrics has antimicrobial efficiency up to 25 washes, whereas the activity of the fabrics containing neem extract remained only up to 10 washes. Azadirachta indica herbal extract nanoparticles were prepared by the coacervation process, followed by crosslinking with glutaraldehyde and application on

Table 1. Antibacterial assessment of nanoparticles of neem extract using the AATCC 100 test method [99].

Fabric treatment	Antimicrobial activity (Bacterial reduction, %)					
	S. aureus	E. coli				
Neem extract	98.73	86.84				
Nano particles of neem extract	100	91.48				

cotton using the traditional pad-dry-cure method (*Table 1*) [99].

The antimicrobial activity of propolis has been investigated since the late 1940s [170]. The propolis extract has been suggested to inhibit the growth of various strains of Gram-positive bacteria, including *Streptococcus* and *Bacillus* [171]. The use of nanopropolis particles has been suggested to show strong efficacy in the range of 85.2 - 100% according to antibiotic susceptibility tests using *Staphylococcus aureus* and *Streptococcal pharyngitis* [172 - 178]. Nanopropolis has been tried clinically [179, 180], but no such trials have been reported in textile applications.

Microencapsulation

Microencapsulation is a process by which droplets of liquid or particles of solid are covered with a continuous film of polymeric material [181 - 183]. This technology has become one of the most promising techniques of imparting functional finishes to textiles. Microencapsulation is more advantageous compared to the conventional processes in terms of economy, energy saving, eco-friendliness and controlled release of substances [184]. However, if the use of a binder during the process of its application to textiles is not carefully controlled, it can affect the handle of the textile. The capsules are applied to fibres as dispersion with a binder using padding, spraying, impregnation, and exhaust or screenprinting techniques [155].

Major interest in the microencapsulation process is currently shown in the application of durable fragrances [65, 185], phase-change materials [186, 187], dyes [118, 188, 189], fire retardants, counterfeiting, antimicrobial agents, and cosmetic textiles [186]. Many techniques have been developed for producing microcapsules, such as phase separation, spray-drying, solvent evaporation as well as interfacial and *in-situ* polymerisation [188, 190 - 196].

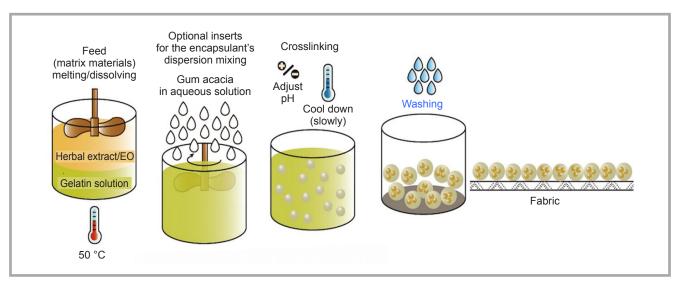


Figure 2. General scheme for preparation of microencapsulation by complex coacervation using gelatin and gum acacia.

Phase separation is a process based on the simultaneous desolvation of oppositely charged polyelectrolyte induced by media modifications. As illustrated in Figure 2, the dispersed phase of dense coacervates made up of concentrated polyelectrolyte and a dilute equilibrium phase is dependent on the pH, ionic strength and polyion concentrations [197 - 199]. Gelatin, a collagen hydrolysis product, is used in complex coacervation. It is associated with different polysaccharides to neutralise opposite charges and thereby form a stable complex. Usually the gelatin is positively charged, and coacervation is induced by anionic colloids [200, 201].

Sathianarayanan et al., [60] used herbal extract as core material and gum acacia as wall material to prepare microcapsules. Such treatment of fabrics has been reported to be durable [202]. Emissive fabrication of chitosan microcapsules encapsulating essential oils in the presence of bio-surfactant has been studied [183, 203 - 205]. This method proved reliable and effective due to the antimicrobial activity of both chitosan and essential oil. Also the prolong bioactivity of the fabric has been suggested to be as a result of the slow release of the antimicrobial agents out of the polymer reservoir [60, 188].

Cross-linking method

Cross-linking happens when a cross-linker makes intermolecular covalent bridges between the polymer chains [13, 206, 207]. Cross-linkers include glutaral-dehyde, genipin, glyoxal, dextran sulfate, 1, 1, 3, 3-tetramethoxypropane, oxidized

cylclodextrins, ethylene glycoldiglycerylether, ethyleneglycoldiglycidylether (EGDE), and diisocyanate [208 - 210]. The approaches to cross-linking are chemical, radiation, and physical methods [115, 211, 212]. Chemical cross-linking happens when the cross-linker makes intermolecular covalent bridges between polymer chains [213, 214]. In radiation cross-linking, heat or a catalyst are not needed, thus no additional toxic chemical is introduced into the system [213, 215]. Radiation polymerisation has been utilizsed by researchers to obtain Interpenetrating Polymer Networks (IPNs) for drug delivery applications, but their applications in textiles have not been reported [216 - 219]. In physical crosslinking, however, a bond is obtained by using cross-linkers that establishe ionic interactions between polymer chains [220]. Chemical cross-linking has really excellent durability with no adverse effect on the tensile strength or handle of the fabric [60]. Physical cross-linking, although not as permanent as chemical cross-linking, possesses good thermal and antimicrobial activity, and is environmentally friendly [60, 221, 222].

To achieve the cross-linking effect of herbal products on fabrics, the herbal extract is mixed with a non-formaldehyde base resin usually with MgCl₂ as catalyst [115]. The fabric is coated by the traditional pad-dry-cure/exhaust process to achieve the cross-linking effect. Joshi et al. [13] used glyoxal as a cross-linking chemical to enhance the application of *Azadirachta indica* extract on polyester/cotton blend fabrics. The antibacterial properties of the fabric were retained after several washing cycles, which clearly demonstrated that carefully selected

resins have the potential to enhance the cross-linking yield of bioactive plant parts irrespective of the substrate. Similarly citric acid was used as a cross-linking agent in the antibacterial finishing of cotton with green tea leaf extracts to increase durability against washing [220]. Also cotton fabrics were treated with two different cross-linking agents, i.e. Butane tetracarboxylic acid (BTCA) and Arcofix NEC (low formaldehyde content). Cotton fabrics treated with the cross linker above showed broad-spectrum antimicrobial activity against Gram-positive and Gram-negative bacteria as well as fungi [3, 223 - 225].

Combination of microencapsulation and cross-linking

Microencapsulation is a physicochemical approach for textile finishing. The synergy of microencapsulation and cross-linking has great potential for the application of natural antimicrobes to textiles [5, 74, 89, 226, 227]. Herbal extracts of pomegranate rind and Ocimum tenuiflorum were used as core materials with acacia gum as the wall material for microencapsulation and non-formadehyde resin as the cross-linking agent [228]. As can be seen from Table 2, cotton fabrics treated using the abovementioned methods showed good and relatively lasting antimicrobial activity compared to the untreated fabrics.

The compound prepared showed excellent antimicrobial activity against both Gram-positive and Gram-negative bacteria [60]. In a similar study conducted by Banupriya et al., [229], it was found that extracts of *Michelia alba* have high antimicrobial efficacy against all types

of Gram-negative bacteria when applied on cotton fabrics using cross-linking and microencapsulation processes. Several other plants have been reported to exhibit excellent bioactive functions [48, 63, 116], some of which have been applied extensively in medicine, but their use in textile applications has not been reported yet due to the lack of suitable methods of application.

Fibre surface modification

Altering the surface properties of fibres is also an interesting way of ensuring the strong adhesion of finishing agents to textiles. Surface modification methods, such as oxygen plasma treatment, ultrasound technology, UV radiation, surface bridging, enzyme treatment and several other techniques have been adopted lately in a quest to impart durable antimicrobial finishes to fabrics using natural products [230 - 233]. Several studies on the use of natural antimicrobial products and colorants combined with these techniques to impart durable finishing treatment have been reported [109, 132, 230, 231, 234].

Chemical modification

Chemical modification is used to impart a deliberate change in the composition or structure of fabrics leading to an improvement in fibre properties [235, 236]. In the pursuit to achieve durable antimicrobial finish with the natural biocide berberine (Figure 3), the surface of cotton fabric was modified with an anionic bridging agent to create electrostatic interactions with cationic natural dye, thereby enhancing the affinity of berberine to cotton fabric [132, 237].

In a similar development, cotton and wool fabrics were modified by pre-treatment with hydrogen peroxide and formic acid to increase the accessible areas [234]. Glyoxal was also used to enhance the application of leaf and seed extract on polyester/cotton blend fabrics [13, 238]. Antibacterial properties were retained after several wash cycles. Also citric acid was used as a cross-linking agent with green tea leaf extracts for antibacterial finishing of cotton fabric. The treatment above was reported to have significantly increased the antibacterial durability of the textile [109, 220].

The use of biological enzymes

Enzyme treatments are used extensively in the textile industry to catalyse chemical reactions due to their effectiveness in improving the wettability, dyeability, bleachability and other finishing processes without fibre devastation [239 - 242]. The biological enzyme proteases, capable of conducting proteolysis, was combined with some natural dyes possessing antimicrobial characteristics and applied on wool fibres [243, 244]. The effects of protease enzyme treatment on dyeing and antimicrobial yield studied demonstrated that the alkaline *protease* enzyme process enhanced the quantity of natural dyes exhausted while maintaining strong antimicrobial activity. This was attributed to the partial or complete damage of the cuticle in the wool, which facilitated easier penetration of natural dye molecules into fibres [243, 245 - 247]. Also antimicrobial peptides form part of a class of naturally occurring antimicrobial molecules which have good antimicrobial activity against a wide range of pathogenic microorganisms. Antimicrobial peptides are short (typically ranging from 12 - 100 amino acid residues in length), exhibit rapid and efficient antimicrobial toxicity against a wide range of pathogens, and constitute critical effector molecules in the innate immune system of both prokaryotic and eukaryotic organisms. Antimicrobial peptides exert their microbicidal effect via the disruption of the microbial cell membrane together with intracellular action [248, 249]. Laverty et al., [249] have done an extensive review of antimicrobial peptides and their mechanism of

The use of plasma technology

The use of physical modification techniques such as low temperature plasma treatment has also been exploited in the application of natural dye/antimicrobial agents [160, 230]. Ghoranneviss et al., [230] studied the dyeing properties of plasma-treated wool fibres with two natural dyes, namely madder and weld. From their findings, they concluded that the treatment of wool fibres with plasma

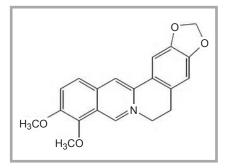


Figure 3. Chemical structure of berberine.

enriches the natural coloration process while enhancing the antimicrobial properties of the fibre. In a similar study conducted by Chen and Chang [231], plasma treatment was suitably introduced as a replacement for metallic mordant treatment, recording impressive results. They used low-temperature microwave plasma to treat cotton fabric prior to grafting with onion skin and onion pulp extracts in order to impart antimicrobial properties to cotton fabrics. They discovered that the functional groups produced on the fibre surface after the low temperature oxygen plasma treatment significantly increased the hydrophilic nature of cotton, thereby enhancing the grafting yield of both the onion skin and pulp extracts [135, 160].

The use of ultra sound and UV technologies

The use of modern fabric coloration technologies such as ultra sound and UV radiation in the application of natural dyes with antimicrobial properties has also been reported [115, 250 - 252]. Wool fibres were dyed with natural dye *lac* (secretion of *Kerria lacca* insect) using both conventional and ultrasonic techniques [232, 253]. In this study, ultrasonic dyeing recorded an impressive 47% increase in lac dye uptake, where the amount of dye extracted was 41% higher than by conventional methods of extraction. In a related study conducted by Kamel et

Table 2. Antimicrobial activity of cotton treated with herbal extracts using various methods [26]; Sa – Staphylococcus aureus , Kp – Klebsiella pneumoniae

	Growth under fabric	Qualitative study zone of inhibition, mm				Qualitative study bacterial growth reduction, %			
		Tulsi extract		Pomegranate extract		Tulsi extract		Pomegranate extract	
		Sa	Кр	Sa	Кр	Sa	Кр	Sa	Кр
Untreated (control)	Present	0	0	0	0	0	0	0	0
Direct treated	Present	12.5	6.6	12.6	7.1	99.9	95.3	99.9	90.8
Micro-encapsulated	Present	10.1	5.8	8.0	5.9	99.9	92.6	99.8	85.2
Cross-linked	Present	9.9	4.9	13.5	6.5	99.2	94.2	98.2	89.2
Micro-encapsulated & cross-linked	Present	10.2	5.4	10.1	8.0	99.8	94.5	99.7	87.1

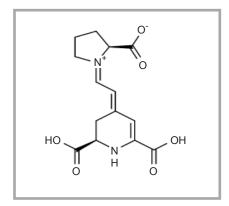


Figure 4. Chemical structure of indicaxanthin.

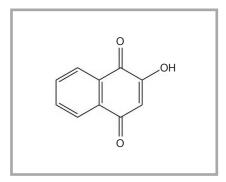


Figure 5. Chemical structure of lawsone.

al., [232] where cationised cotton fabric was dyed using conventional and ultrasonic methods, the ultrasonic dyeing technique recorded a 66.5% improvement in lac dye uptake compared with the conventional method. Also the antimicrobial property of the ultrasonic dyeing technique was much better than that of the conventional method. In a similar comparative study of the two processes conducted by Vankar et al., [254] using dye extract of eclipta prostrata, cotton fabric was dyed using both conventional and sonicator dyeing. The dyeing efficiency of sonicator dyeing was much higher than in the conventional method.

In a recent study conducted by Guesmi et al., [255] an assessment was made of the dyeing properties of modified acrylic fibres dyed with *indicaxanthin* (*Figure 4*) using both conventional and sonicator dyeing.

In line with results obtained in the previous venture, sonicator dyeing recorded higher dye uptake, better wash and light fastness, an excellent antimicrobial property with a short process time, and energy consumption compared with the conventional dyeing technique. A similar result was obtained by Vankar et al., [233]

when they dyed cotton with *Symplocos spicata* plant dye extract using the two technologies.

Iqbal et al., [256] studied the effect of UV irradiation on henna-dyed cotton fabrics. In their research, henna and cotton were UV irradiated separately and then coloured. A second set of henna and cotton were not irradiated under UV light. An investigation into the properties of the two samples revealed that the colour strength and shade of fabrics dyed with irradiated henna were much higher compared to those of non-irradiated henna samples, which was attributed to the hydrolytic degradation of lawsone (the colouring matter in henna, see Figure 5) upon irradiation, resulting in a considerable increase in the sorption of the coloured matter.

They further concluded that the UV irradiation of both henna dye and cotton is necessary for the attainment of an excellent colour strength and antimicrobial property [109, 256]

Future prospects

Consumers are now increasingly aware of a hygienic lifestyle, hence it is imperative that textile products should be finished with safe and effective antimicrobial agents. With the enhanced consumer awareness of eco-safety, research has now focused on the use of sustainable and environmentally friendly materials. In recent years, considerable attention has been given to products produced from non-food crops for use in the textile industry [13]. Based on the biocompatibility, biodegradability and non-toxicity of these natural products, in addition to their recently discovered properties such as insect repellency, deodorisation, flame retardancy, UV protection, and antimicrobial activity, these natural products are gaining popularity around the world for producing more appealing and highly functional value-added textiles.

The focus on plant derived antimicrobial agents with potent antimicrobial activities and their application to textiles has increased significantly considering the number of research publications on the topic over the past few years. The contribution of plant-based antimicrobial agents to green nanotechnology in recent years for the development of bioactive textiles has become a reality.

The demand for antimicrobial textiles will be spurred primarily by the growing awareness of consumers of the importance of personal hygiene and the health risk posed by microorganisms.

Considering the level of research into so-called 'green technology' today, it is possible that antimicrobial finishing of textiles may be tagged as naturally or synthetically protected, which will offer consumers a greater choice in their quest to satisfy their health needs with respect to textiles.

Reference

- Lacasse K and Baumann W. Textile Chemicals: Environmental Data and Facts. Germany: Springer-Verlag Berlin Heidelberg; 2004.
- Blackburn RS. Sustainable Textiles:Life Cycle and Environmental Impact. Uk: Woodhead Publishing Limited; 2009.
- Shahidi S and Jakub W. Antibacterial Agents in Textile Industry. *International Journal of Textile Science*. 2013; 4(3): 387-406
- Gao Y and Cranston R. Recent advances in antimicrobial treatments of textiles. Textile Research Journal; 78(1): 60-72.
- Purwar R and Joshi M. Recent Developments in Antimicrobial Finishing of Textiles — A Review. AATCC Review, 2004; 4: 22–26.
- Simoncic B and Tomsic B. Structures of Novel Antimicrobial Agents for Textiles

 A Review. Textile Research Journal.

 2010; 80(16): 1721-1737.
- Li R, Hu P, Ren X, Worley SD and Huang TS. Antimicrobial N-halamine modified chitosan films. *Carbohydrate* polymers. 2013; 92(1): 534-539.
- Nichifor M, Constantin M, Mocanu G, Fundueanu G, Branisteanu D and Costuleanu M. New multifunctional textile biomaterials for the treatment of leg venous insufficiency. J Mater Sci: Mater Med. 2009; 20(4): 975-982.
- Demand for antimicrobial fibres, textiles and apparel is set for strong growth: Performance Apparel Market [press release], R. Anson Munich, Germany, 2014, p. 1.
- Isabella Gouveia C. Current Research, Technology and Education Topics in Applied Microbiology and Microbial Biotechnology. In: Ed. AM-V, editor. Formatex; Spain 2010, p. 407-414.
- Zinner SH. Overview of antibiotic use and resistance: setting the stage for tigecycline. Clinical infectious diseases: an official publication of the Infectious Diseases Society of America, 2005; 41 Suppl 5: S289-S292.
- Mathew BP, Kumar A, Sharma S, Shukla PK and Nath M. An eco-friendly synthesis and antimicrobial activities of

- dihydro-2H-benzo- and naphtho-1,3-oxazine derivatives. *European Journal of Medicinal Chemistry*, 2010; 45(4): 1502-1507.
- Joshi M, Purwar R, Wazed Ali S and Rajendran S. Antimicrobial Textiles for Health and Hygiene Applications Based on Eco-Friendly Natural Products. In: Anand SC, Kennedy JF, Miraftab M, Rajendran S, editors. Medical and Healthcare Textiles: Woodhead Publishing; 2010. p. 84-92.
- Shahid ul I, Shahid M and Mohammad F. Perspectives for natural product based agents derived from industrial plants in textile applications a review. Journal of Cleaner Production, 2013; 57(0): 2-18.
- El-Shafei A, ElShemy M and Abou-Okeil A. Eco-friendly finishing agent for cotton fabrics to improve flame retardant and antibacterial properties. *Carbo*hydrate polymers, 2015; 118(0): 83-90.
- Prasannabalaji NMG, Sivanandan RN and Kumaran S. Pugazhvendan SR Antibacterial activities of some Indian traditional plant extracts. Asian Pacific Journal of Tropical Disease, 2012: 291-295.
- Mahesh S, Manjunatha AH, Reddy V and Kumar G. Studies on Antimicrobial Textile Finish Using Certain Plant Natural Products. International Conference on Advances in Biotechnology and Pharmaceutical Sciences (ICABPS'2011) Bangkok Dec. 2011.
- Hussain AI, Anwar F, Chatha SAS, Latif S, Sherazi STH and Ahmad A. Chemical composition and bioactivity studies of the essential oils from two Thymus species from the Pakistani flora. LWT - Food Science and Technology, 2013; 50(1): 185-192.
- Mangalanayaki N and Niroshi R. Antibacterial Activity of Leaves and Stem Extract of Carica papaya L. International journal of advances in pharmacy, biology and chemistry, 2013; 2(3): 233-238
- Hooda S, Khambra K, Yadav N and Sikka VK. Eco-friendly Antimicrobial Finish for Wool Fabric. *J Life Sci*, 2013; 5(1): 11-16.
- Gobalakrishnan R, Kulandaivelu M, Bhuvaneswari R, Kandavel D and Kannan L. Screening of wild plant species for antibacterial activity and phytochemical analysis of Tragia involucrata L. Journal of Pharmaceutical Analysis, 2013; 3(6): 460-465.
- Bauer C, Buchgeister J, Hischier R, Poganietz WR, Schebek L and Warsen J. Towards a framework for life cycle thinking in the assessment of nanotechnology. J Cleaner Prod., 2008; 16(910): 26-32.
- Burnett-Boothroyd SC and McCarthy BJ. Antimicrobial treatments of textiles for hygiene and infection control applications. an industrial perspective, 2011

- ed: Woodhead Publishing Limited; 2011. p. 196-209.
- Elsner P. Antimicrobials and the Skin Physiological and Pathological Flora. Biofunctional Textiles and the Skin, 2006: 35-41.
- Gopalakrishnan D, Aswini, RK. Antimicrobial Finishes 2013 [cited 2014 14th May]. Available from: http://www.fibre-2fashion.com/industry-article/pdffiles/antimicrobial-finishes.pdf.
- Elshafei A and El-Zanfaly HT. Application of Antimicrobials in the Development of Textiles. Asian Journal of Applied Sciences, 2011; 4(6): 585-595.
- Smith EJ, Williams JT, Walsh SE and Painter P. Comparison of Antimicrobial Textile Treatments. In: Anand SC, Kennedy JF, Miraftab M, Rajendran S, editors. Medical and Healthcare Textiles: Woodhead Publishing; 2010. p. 38-47.
- Faheem U. Environmental Concerns in Antimicrobial Finishing of Textiles. International Journal of Textile Science. 2014; 3(1A): 15-20.
- Pedrosa M, Granadeiro ML, Piskin E, Henriques M and Gouveia IC. Novel Bioactive Textile -Base Materials. Mechanism of Action and Potential Antimicrobial Properties. Proceedings of Global Engineering, Science and Technology Conference, 2013; 987: 2069-2072.
- Barbara Filipowska ER, Anetta W and Edyta M-Z. New Method for the Antibacterial and Antifungal Modification of Silver Finished Textiles. Fibres & Textiles in Eastern Europe, 2011; 4(87): 124-128.
- 31. Gyawali R and Ibrahim SA. Natural products as antimicrobial agents. *Food Control*. 2014; 46(0): 412-429.
- García A, Bocanegra-García V, Palma-Nicolás JP and Rivera G. Recent advances in antitubercular natural products. European Journal of Medicinal Chemistry, 2012; 49(0): 1-23.
- Dembitsky VM. Naturally occurring bioactive Cyclobutane-containing (CBC) alkaloids in fungi, fungal endophytes, and plants. *Phytomedicine*, 2014; 21(12): 1559-1581.
- Yoneyama K, Natsume M. Allelochemicals for Plant–Plant and Plant–Microbe Interactions. Reference Module in Chemistry, Molecular Sciences and Chemical Engineering, 2013; 20(16): 234-239
- Awouafack MD, Tane P, Kuete V, Eloff JN - Sesquiterpenes from the Medicinal Plants of Africa. In: Kuete V, editor. Medicinal Plant Research in Africa. Oxford: Elsevier, 2013. p. 33-103.
- Rifai S, Fassouane A, El-Abbouyi A, Wardani A, Kijjoa A, Van Soest R. Screening of antimicrobial activity of marine sponge extracts. *Journal of Medical Mycology*. 2005; 15(1): 33-38.
- Rakholiya KD, Kaneria MJ, Chanda SV. Chapter 11 - Medicinal Plants as Alternative Sources of Therapeutics against Multidrug-Resistant Pathogenic Micro-

- organisms Based on Their Antimicrobial Potential and Synergistic Properties. In: Kon MKRV, editor. Fighting Multidrug Resistance with Herbal Extracts, Essential Oils and their Components. San Diego: Academic Press; 2013. p. 165-179.
- 38. Kenny O, Smyth TJ, Walsh D, Kelleher CT, Hewage CM, Brunton NP. Investigating the potential of under-utilised plants from the Asteraceae family as a source of natural antimicrobial and antioxidant extracts. *Food Chemistry*, 2014;161(0): 79-86.
- Hübsch Z, Van Zyl RL, Cock IE, Van Vuuren SF. Interactive antimicrobial and toxicity profiles of conventional antimicrobials with Southern African medicinal plants. South African Journal of Botany, 2014; 93(0): 185-197.
- Samanta AK, Agarwal, P. Application of natural dyes on textiles. *Indian J Fibre* Text Res. 2009; 34: 384-389.
- Elisa Friska Romasi JK, Adolf Jan Nexson Parhusip. Antibacterial activity of papaya leaf extracts against pathogenic bacteria. *Makara teknologi*, 2011; 15(2): 173-177.
- Ogunkunle J, Tonia AL. Ethnobotanical and phytochemical studies on some species of Senna in Nigeria. *Afr. J Bio*technol., 2006; 5(21): 2020–2023.
- 43. Okunola A, Muyideen TH, Chinedu P. Anokwuru, Tomisin J, Harrison A, Victor UO, Babatunde EE. Comparative studies on antimicrobial properties of extracts of fresh and dried leaves of Carica papaya (L) on clinical bacterial and fungal isolates. Advances in Applied Science Research, 2012; 3(5): 3107-3114.
- Luo X, Pires D, Aínsa JA, Gracia B, Mulhovo S, Duarte A. Antimycobacterial evaluation and preliminary phytochemical investigation of selected medicinal plants traditionally used in Mozambique. *Journal of Ethnopharmacology*, 2011: 137(1): 114-120.
- Mansoor TA, Borralho PM, Luo X, Mulhovo S, Rodrigues CMP, Ferreira M-JU. Apoptosis inducing activity of benzophenanthridine-type alkaloids and 2-arylbenzofuran neolignans in HCT116 colon carcinoma cells. *Phytomedicine*, 2013; 20(10): 923-929.
- 46. Baskaran C, Bai VR, Velu S, Kumaran K. The efficacy of Carica papaya leaf extract on some bacterial and a fungal strain by well diffusion method. Asian Pacific Journal of Tropical Disease. 2012; Supplement 2(0): S658-S662.
- 47. Juárez-Rojop IE, Tovilla-Zárate CA, Aguilar-Domínguez DE, Fuente LFRdl, Lobato-García CE, Blé-Castillo JL, Phytochemical screening and hypoglycemic activity of Carica papaya leaf in streptozotocin-induced diabetic rats. Revista Brasileira de Farmacognosia, 2014; 24(3): 341-347.
- 48. Vij T, Prashar Y. A review on medicinal properties of Carica papaya Linn. *Asian*

- Pacific Journal of Tropical Disease, 2015; 5(1): 1-6.
- Chanda S, Dudhatra S, Kaneria M. Antioxidative and antibacterial effects of seeds and fruit rind of nutraceutical plants belonging to the Fabaceae family. Food & function, 2010; 1(3): 308-315.
- Gutarowska B, Machnowski W, Kowzowicz Ł. Antimicrobial activity of textiles with selected dyes and finishing agents used in the textile industry. *Fibers and Polymers*, 2013; 14(3): 415-422.
- PerumalSamy R. GP, Houghton P. Purification of antibacterial agents from Tragiainvolucrata: a popular tribal medicine for wound healing. *J Ethnopharmacol*, 2006; 107(1): 99–106.
- Wang D, Huang L, Chen S. Senecio scandens Buch.-Ham.: A review on its ethnopharmacology, phytochemistry, pharmacology, and toxicity. *Journal* of Ethnopharmacology, 2013; 149(1): 1-23.
- Ye Y, Li X-Q, Tang C-P, Yao S. Natural products chemistry research: progress in China in 2011. *Chinese Journal of Natural Medicines*, 2013; 11(2): 97-109.
- Zhang H-F, Yang X-H, Wang Y. Microwave assisted extraction of secondary metabolites from plants: Current status and future directions. *Trends in Food Science & Technology*, 2011; 22(12): 672-688.
- 55. Nikolić M, Glamočlija J, Ferreira ICFR, Calhelha RC, Fernandes Â, Marković T, Chemical composition, antimicrobial, antioxidant and antitumor activity of Thymus serpyllum L., Thymus algeriensis Boiss. and Reut and Thymus vulgaris L. essential oils. *Industrial Crops* and Products, 2014; 52(0): 183-190.
- Ouariachi EE, Hamdani I, Bouyanzer A, Hammouti B, Majidi L, Costa J. Chemical composition and antioxidant activity of essential oils of Thymus broussonetii Boiss. and Thymus algeriensis Boiss. from Morocco. Asian Pacific Journal of Tropical Disease, 2014; 4(4): 281-286.
- Zouari N, Fakhfakh N, Zouari S, Bougatef A, Karray A, Neffati M. Chemical composition, angiotensin I-converting enzyme inhibitory, antioxidant and antimicrobial activities of essential oil of Tunisian Thymus algeriensis Boiss. et Reut. (Lamiaceae). Food and Bioproducts Processing, 2011; 89(4): 257-265.
- Nejad Ebrahimi S, Hadian J, Mirjalili MH, Sonboli A, Yousefzadi M. Essential oil composition and antibacterial activity of Thymus caramanicus at different phenological stages. Food Chemistry, 2008; 110(4): 927-931.
- Deepti Guptaa AL. cotton treated with natural antimicrobial product. *Indian Journal of Fibre & Textile Research*, 2007; 32: 88-92.
- Sathianarayanan MP Bhat NV, Kokate SS, Walunj VE. Antibacterial finish for cotton fabric from herbal products. *Indi-*

- an Journal of Fibre & Textile Research, 2010; 35: 50-58.
- Yoon JI, Bajpai VK, Kang SC. Synergistic effect of nisin and cone essential oil of MetasequoiaglyptostroboidesMiki ex Hu against Listeria monocytogenesin milk samples. Food Chem Toxicol., 2011; 49: 109-114.
- Yang VW, Clausen CA. Antifungal effect of essential oils on southern yellow pine. *International Biodeterioration*& *Biodegradation*, 2007; 59: 302-306.
- Vinoth B. Manivasagaperumal R, Balamurugan S. Phytochemical analysis and antibacterial activity of Moringaoleifera Lam. *Int J Res Biol Sci.* 2012; 2(3): 98-102.
- Buckle J. Clinical aromatherapy and AIDS. J Assoc Nurse Aids Care, 2002; 13: 81-90.
- Wang CX, Chen SL. Aromachology and its application in the textile field. *Fibres* & *Textile in Eastern Europe*, 2005; 13, 6(54); 41-44.
- Walentowska J, Foksowicz-Flaczyk J. Thyme essential oil for antimicrobial protection of natural textiles. *Interna*tional Biodeterioration & Biodegradation, 2013; 84: 407-411.
- Giordani R, Regli P, Kaloustian J, Mikail C, Abou L, Portugal H. Antifungal effect of various essential oils against Candidaalbicans. Potentiation of antifungal action of amphotericin B by essential oil from Thymus vulgaris. *Phytother Res.*, 2004;18: 990–995.
- Yamazaki K, Yamamoto T, Kawai Y, Inoue N. Enhancement of antilisterial activity of essential oil constituents by nisin and diglycerol fatty acid ester. Food Microbiol., 2004; 21: 283–289.
- Shahverdi AR, Rafi F, Fazeli MR, Jamalifar H. Enhancement of antimicrobial activity of furazolidone and nitrofurantoin against clinical isolates of Enterobacteriaceae by piperitone. *Int J. Aromather*, 2004; 14: 77-80.
- Rajkovic A, Uyttendaele M, Courtens T, Debevere. Antimicrobial effect of nisin and carvacrol and competition between Bacillus cereus and Bacillus circulansin acuum-packed potato puree. *J. Food Microbiol.*, 2005; 22: 189–197.
- Pyun MS, Shin S. Antifungal effects of the volatile oils from Allium plants against Trichophytonspecies and synergism of the oils with ketaconazole. *Phytomedicine*, 2006; 13: 394–400.
- Grande MJ, Lopez R, Abriouel H, Valdivia E, Ben ON, Maqueda M, Martinez-Canamero M, Galvez A. Treatment of vegetable sauces with enterocinAS-48 alone or in combination with phenolic compounds to inhibit proliferation of Staphylococcus aureus. *J. Food Prot.*, 2007; 70: 405–411.
- Rosato A, Vitali C, de Laurentis N, Armenise D, Nulillo MA. Antibacterial effect of some essential oils administered alone or in combination with norfloxacin. *Phytomedicine*, 2007;14: 727–732.

- Dimitrijevic SI, Mihajloski KR, Antonovic DG, Milanovic-Stevanovic MR, Mijin, DZ. A study of the synergistic antilisterial effects of a sub-lethal dose of lactic acid and essential oils from Thymus vulgaris L., Rosmarinusofficinalis L. and Origanumvulgare L. Food Chem Toxicol., 2007; 104: 774-782
- 75. Moosavy MH, Basti AA, Misaghi A, Salehi TZ, Abbasifar R, Ebrahimzadeh Mousavi HA, Alipour M, Razav NE, Gandomi H, Noori N. Effect of Zataria-multifloraBoiss. Essential oil and nisin on Salmonella typhimuriumand Staphylococcus aureusin a food model system and on the bacterial cell membranes. Food Res Int., 2008; 41: 1050-1057.
- Yukie H, Ogita A, Tanaka T, Fujita K. Involvement of inhibition of chitin synthase activity in anethole-induced morphological changes of filamentous fungus Mucor mucedo. *Journal of Bio*technology, 2008;136, Supplement(0): S739.
- 77. Vieira PRN, de Morais SM, Bezerra FHQ, Travassos Ferreira PA, Oliveira ÍR, Silva MGV. Chemical composition and antifungal activity of essential oils from Ocimum species. *Industrial Crops and Products*, 2014; 55(0): 267-271.
- Kang P, Kim KY, Lee HS, Min SS, Seol GH. Anti-inflammatory effects of anethole in lipopolysaccharide-induced acute lung injury in mice. *Life Sciences*, 2013; 93(24): 955-961.
- Delgado B, Fernandez PS, Palop A, Periago PM. Effect of thymol and cymene on Bacillus cereus vegetative cells evaluated through the use of frequency distribution. Food Microbiol., 2004; 21: 327-334.
- Burt SA, Van der zee R, Koets AP, de Graaff AM, van Knapen F, Gaastra W, Haagsman HP, Veldhuizen EJ. Carvacrol induces heat shock protein 60 and inhibits synthesis of flagellin in Escherichia coli O157:H7. Appl Environ Microbiol., 2007; 73: 4484–4490.
- Trombetta D, Castelli F, Sarpietro MG, Venuti V, Cristani M, Daniele C, Saija A, Mazzanti G, Bisignano G. Mechanisms of antibacterial activity of three monoterpenes. *Agents Chemother*, 2005; 49: 2474–2478.
- Hayouni E, Bouix M, Abedrabba M, Leveau JY, Hamdi M. Mechanism of action of Melaleucaarmillaris(Sol. Ex Gaertu) Sm. essential oil on six LAB strains as assessed by multiparametric flow cytometry and automated microtiter-based assay. Food Chem., 2008; 111: 707-718.
- Pandima DK, ArifNisha S, Sakthivel R, KaruthaPandian S. Eugenol (an essential oil of clove) acts as an antibacterial agent against Salmonella typhi by disrupting the cellular membrane. *J. Eth-nopharmacol.*, 2010;130: 107-115.
- 84. Santiesteban-Lopez A, Palour E, López-Malo A. Susceptibility of foodborne bacteria to binary combinations

- of antimicrobials at selected a(w) and pH. *J. Appl Microbiol.*, 2007; 102: 486–497
- Haney EF, Petersen AP, Lau CK, Jing W, Storey DG, Vogel HJ. Mechanism of action of puroindoline derived tryptophan-rich antimicrobial peptides. Biochimica et Biophysica Acta (BBA) Biomembranes, 2013; 1828(8): 1802-1813.
- Manzini MC, Perez KR, Riske KA, Bozelli Jr JC, Santos TL, da Silva MA, Peptide:lipid ratio and membrane surface charge determine the mechanism of action of the antimicrobial peptide BP100. Conformational and functional studies. *Biochimica et Biophysica Acta (BBA) Biomembranes*, 2014;1838(7): 1985-1999.
- Wang P, Bang J-K, Kim HJ, Kim J-K, Kim Y, Shin SY. Antimicrobial specificity and mechanism of action of disulfideremoved linear analogs of the plantderived Cys-rich antimicrobial peptide Ib-AMP1. *Peptides*. 2009; 30(12): 2144-2149.
- Liu H, Pei H, Han Z, Feng G, Li D.
 The antimicrobial effects and syner-gistic antibacterial mechanism of the combination of ε-Polylysine and nisin against Bacillus subtilis. Food Control, 2015;47(0): 444-4450.
- 89. de Azeredo GA, Stamford TLM, Nunes PC, Gomes Neto NJ, de Oliveira MEG, de Souza EL. Combined application of essential oils from Origanum vulgare L. and Rosmarinus officinalis L. to inhibit bacteria and autochthonous microflora associated with minimally processed vegetables. Food Research International, 2011; 44(5): 1541-1548.
- Valero M, Francés E. Synergistic bactericidal effect of carvacrol, cinnamal-dehyde or thymol and refrigeration to inhibit Bacillus cereus in carrot broth. Food Microbiology, 2006; 23(1): 68-73.
- Valero M, Giner MJ. Effects of antimicrobial components of essential oils on growth of Bacillus cereus INRA L2104 in and the sensory qualities of carrot broth. *International journal of food microbiology*, 2006; 106(1): 90-94.
- Pei RS, Zhou F, Ji BP, Xu J. Evaluation of combined antibacterial effects of eugenol, cinnamaldehyde, thymol, and carvacrol against E. coli with an improved method. *Journal of food science*, 2009; 74(7): M379-M383.
- Turgis M, Vu KD, Dupont C, Lacroix M. Combined antimicrobial effect of essential oils and bacteriocins against foodborne pathogens and food spoilage bacteria. Food Research International. 2012; 48(2): 696-702.
- 94. Rosato A, Vitali C, Piarulli M, Mazzotta M, Argentieri MP, Mallamaci R. In vitro synergic efficacy of the combination of Nystatin with the essential oils of Origanum vulgare and Pelargonium graveolens against some Candida species. *Phytomedicine*, 2009; 16(10): 972-975.

- Rosato A, Vitali C, De Laurentis N, Armenise D, Antonietta Milillo M. Antibacterial effect of some essential oils administered alone or in combination with Norfloxacin. *Phytomedicine*, 2007; 14(11): 727-732.
- Fadli M, Saad A, Sayadi S, Chevalier J, Mezrioui N-E, Pagès J-M. Antibacterial activity of Thymus maroccanus and Thymus broussonetii essential oils against nosocomial infection bacteria and their synergistic potential with antibiotics. *Phytomedicine*, 2012; 19(5): 464-471.
- 97. Matan N, Woraprayote W, Saengkrajang W, Sirisombat N. Durability of rubberwood (Heveabrasiliensis) treated with peppermint oil, eucalyptus oil, and their main components. *International Biodeterioration& Biodegradation*, 2009; 63: 621-625.
- El-Tahlawy KF, Magda A. El-Bendary, Adel G. Elhendawy, and Samuel M. Hudson. The antimicrobial activity of cotton fabrics treated with different crosslinking agents and chitosan. Carbohydrate polymers, 2005; 60(4): 421-430.
- Hasabo MA, Rajendran R, Balakumar C. Nanoherbal coating of cotton fabric to enhance antimicrobial durability. *Elix-ir Appl Chem.*, 2012; 45: 7840-7843.
- 100. Jeyakumar E, Lawrence R, Pal T. Comparative evaluation in the efficacy of peppermint (Mentha piperita) oil with standards antibiotics against selected bacterial pathogens. Asian Pacific Journal of Tropical Biomedicine, 2011; 1(2, Supplement): S253-S257.
- 101. Khare AK, Biswas AK, Sahoo J. Comparison study of chitosan, EDTA, eugenol and peppermint oil for antioxidant and antimicrobial potentials in chicken noodles and their effect on colour and oxidative stability at ambient temperature storage. Food Science and Technology, 2014; 55(1): 286-293.
- 102. Tarek N, Hassan HM, AbdelGhani SMM, Radwan IA, Hammouda O, El-Gendy AO. Comparative chemical and antimicrobial study of nine essential oils obtained from medicinal plants growing in Egypt. Beni-Suef University Journal of Basic and Applied Sciences, 2014; 3(2): 149-156.
- 103. Riachi LG, De Maria CAB. Peppermint antioxidants revisited. Food Chemistry, 2015; 176(0): 72-81.
- 104. Misaghi A, Basti AA. Effects of ZatariamultifloraBoiss. Essential oil and nisin on Bacillus cereus ATCC 11778. Food Control, 2007; 18: 1043–1049.
- 105. Rodriguez A, Batlle R, Nerin C.The use of natural essential oils as anti- microbial solutions in paper packaging. Part II. Progress in Organic Coatings. 2007; 60: 33-38.
- 106. Bassole IH, Juliani HR. Essential oils in combination and their antimicrobial properties. *Molecules*, 2012;17(4): 3989-4006.

- 107. Walentowska J, Foksowicz-Flaczyk J. Thyme essential oil for antimicrobial protection of natural textiles. *Interna*tional Biodeterioration & Biodegradation. 2013; 84(0): 407-411.
- 108. Mann A, Amupitan JO, Oyewale AO. Antibacterial activity of terpenoidal fractions from Anogeissusleiocarpus and Terminaliaavicennioides against community acquired infections. *Afr J Pharm Pharmacol.*, 2009; 3(1): 22-25.
- 109. Masoud B. Kasiri SS. Natural dyes and antimicrobials for green treatment of textiles. *Environmental Chemistry Letters*, 2014;12(1): 1-13.
- 110. Shahmoradi Ghaheh F, Mortazavi SM, Alihosseini F, Fassihi A, Shams Nateri A, Abedi D. Assessment of antibacterial activity of wool fabrics dyed with natural dyes. *Journal of Cleaner Production*, 2014; 72(0): 139-45.
- 111. Hayashi MA, Bizerra FC, Da Silva PI. Antimicrobial compounds from natural sources. *Front Microbiol*. 2013;4-9
- 112. Sivakumar V, Vijaeeswarri J, Anna JL. Effective natural dye extraction from different plant materials using ultrasound. *Industrial Crops Prod.*, 2011; 33: 116–122.
- 113. Prusty AK, Das T, Nayak A, Das NB. Colourimetric analysis and antimicrobial study of natural dyes and dyed silk. *Journal of Cleaner Production*. 2010; 18(16–17): 1750-1756.
- 114. Baliarsingh S, Jena J, Das T, Das NB. Role of cationic and anionic surfactants in textile dyeing with natural dyes extracted from waste plant materials and their potential antimicrobial properties. *Industrial Crops and Products*, 2013; 50(0): 618-624.
- 115. Shahid M, Shahid ul I, Mohammad F. Recent advancements in natural dye applications: a review. *Journal of Cleaner Production*, 2013; 53(0): 310-331
- 116. Singh R, Jain A, Panwar S, Gupta D, Khare SK. Antimicrobial activity of some natural dyes. *Dyes and Pig*ments, 2005; 66(2): 99-102.
- 117. Dawson TL. Biosynthesis and synthesis of natural colours. *Color Technol.*, 2009; 25: 61–73.
- 118. Chakraborty JN. Dyeing with natural dyes. In: Chakraborty JN, editor. Fundamentals and Practices in Colouration of Textiles: Woodhead Publishing India; 2014. p. 233-261.
- 119. Zhang B, Wang L, Luo L, King MW. Natural dye extracted from Chinese gall the application of color and antibacterial activity to wool fabric. *Journal of Cleaner Production*. 2014; 80(0): 204-210.
- 120. Ayfer C, Gökçen Y, Çelik HK. Antimicrobial effect of natural dyes on some pathogenic bacteria. *African Journal of Biotechnology*, 2009; 8(2): 291-293.
- 121. Mirjalili M, Karimi L. Antibacterial dyeing of polyamide using turmeric as a natu-

- ral dye. Autex Research Journal. 2014; 13(2): 51-56.
- 122. Jafari S, Izadan H, Khoddami A, Zarrebini M. Investigation into the Dyeing of Soybean Fibres with Natural Dyes and their antimicrobial properties. *J Prog Color, Colorants, Coatings*, 2014; 7: 2012-2018.
- 123. Kulkarni SS, Gogkhale AV, Bodake UM, Pathade GR. Cotton dyeing with natural dye extracted from pomegranate (Punica granatum) peel. *Universal J Environ* Res Technol., 2011; 1(12): 135-139.
- 124. Silva LM, Hill LE, Figueiredo E, Gomes CL. Delivery of phytochemicals of tropical fruit by-products using poly (dl-lactide-co-glycolide) (PLGA) nanoparticles: Synthesis, characterization, and antimicrobial activity. Food Chemistry, 2014; 165(0): 362-370.
- 125. Mariem C, Sameh M, Nadhem S, Soumaya Z, Najiba Z, Raoudha EG. Antioxidant and antimicrobial properties of the extracts from Nitraria retusa fruits and their applications to meat product preservation. *Industrial Crops and Products*, 2014; 55(0): 295-303.
- 126. Bag A, Bhattacharyya SK, Pal NK, Chattopadhyay RR. In vitro antimicrobial potential of Terminalia chebula fruit extracts against multidrug-resistant uropathogens. Asian Pacific Journal of Tropical Biomedicine, 2012; 2(3, Supplement): S1883-S1887.
- 127. Akhavan M, Jahangiri S, Shafaghat A. Studies on the antioxidant and antimicrobial activity and flavonoid derivatives from the fruit of Trigonosciadium brachytaenium (Boiss.) Alava. *Industrial Crops and Product*, 2015; 63(0): 114-118.
- 128. Al-Zoreky NS. Antimicrobial activity of pomegranate (Punica granatum L.) fruit peels. *International journal of food microbiology*, 2009; 134(3): 244-248.
- 129. Mphahlele RR, Fawole OA, Stander MA, Opara UL. Preharvest and post-harvest factors influencing bioactive compounds in pomegranate (Punica granatum L.) A review. *Scientia Horticulturae*, 2014; 178(0): 114-123.
- 130. Hleba L, Vuković N, Horská E, Petrová J, Sukdolak S, Kačániová M. Phenolic profile and antimicrobial activities to selected microorganisms of some wild medical plant from Slovakia. Asian Pacific Journal of Tropical Disease, 2014; 4(4): 269-274.
- 131. De Santis D, Moresi M, Gallo AM., Petruccioli M. Assessment of the dyeing properties of pigments from Monascus purpureus. 80:1072–1079. J Chem Technol Biotechnol., 2005; 80: 1072-1079.
- 132. Kim S, Cha M, Oh ET, Kang S, So J, Kwon Y. Use of protease produced by Bacillus sp. SJ-121 for improvement of dyeing quality in wool and silk. *Bio*technol Bioprocess Eng., 2005; 10: 186-191.

- 133. Velmurugan P, Lee YH, Venil CK, Lakshmanaperumalsamy P, Chae J-C, Oh B-T. Effect of light on growth, intracellular and extracellular pigment production by five pigment-producing filamentous fungi in synthetic medium. *Journal of Bioscience and Bioengineering*, 2010; 109(4): 346-350.
- 134. Houbraken J, de Vries RP, Samson RA. Chapter Four - Modern Taxonomy of Biotechnologically Important Aspergillus and Penicillium Species. In: Sima S, Geoffrey MG, editors. Advances in Applied Microbiology, Volume 86: Academic Press; 2014. p. 199-249.
- 135. Jiang Y-h, Jiang X-I, Wang P, Mou H-j, Hu X-k, Liu S-q. The antitumor and antioxidative activities of polysaccharides isolated from Isaria farinosa B05. *Mi*crobiological Research, 2008; 163(4): 424-430.
- 136. Velmurugan PCJ, Lakshmanaperumalsamy P, Yun B, Lee K, Oh B. Assessment of the dyeing properties of pigments from five fungi and anti-bacterial activity of dyed cotton fabric and leather. Color Technol., 2010; 125: 334-341.
- 137. Vaidyanathan J B-LZ, Adivarekar R.V, Nerurkar M. Production, partial characterization, and use of a red biochrome produced by Serratia sakuensis subsp. nov strain KRED for dyeing natural fibers. Appl Biochem Biotechnol., 2012; 166: 321-335.
- 138. Sharma D, Gupta C, Aggarwal S, Nagpal N. Pigment extraction from fungus for textile dyeing. *Indian Journal of Fibre & Textile Research*, 2012; 37: 68–73.
- 139. Zhang Y, Wang Z, Zhang J, Chen C, Wu Q, Zhang L. Quantitative determination of chitinolytic activity of lysozyme using half-deacetylated chitosan as a substrate. *Carbohydrate polymers*, 2011; 85(3): 554-549.
- 140. Taghizadeh SM, Davari G. Preparation, characterization, and swelling behavior of N-acetylated and deacetylated chitosans. Carbohydrate polymers, 2006; 64(1): 9-15.
- 141. Aklog YF, Dutta AK, Izawa H, Morimoto M, Saimoto H, Ifuku S. Preparation of chitosan nanofibers from completely deacetylated chitosan powder by a downsizing process. *International Journal of Biological Macromolecules*, 2015; 72(0): 1191-1195.
- 142. Lou M-M, Zhu B, Muhammad I, Li B, Xie G-L, Wang Y-L. Antibacterial activity and mechanism of action of chitosan solutions against apricot fruit rot pathogen Burkholderia seminalis. Carbohydrate Research, 2011; 346(11): 1294-1301.
- 143. Li L, Wang L, Li J, Jiang S, Wang Y, Zhang X, et al. Insights into the mechanisms of chitosan–anionic polymers-based matrix tablets for extended drug release. *International Journal of Pharmaceutics*, 2014; 476(1–2): 253-265.

- 144. Kong M, Chen XG, Xing K, Park HJ. Antimicrobial properties of chitosan and mode of action: A state of the art review. *International journal of food microbiology*, 2010; 144(1): 51-63.
- 145. Amada Y E-L, Alain D, Carlos AV, Francisco MG, Waldo MA-M, Physical properties and antibacterial activity of chitosan/acemannan mixed systems. Carbohydrate Polymers, 2015, 115: 707-714
- 146. Eugene Khor. Chitin and Chitosan Tissue Engineering and Stem Cell Research, In Chitin (Second Edition), edited by Eugene Khor, Elsevier, Oxford, 2014, P. 51-66,
- 147. Chung Y, Tsai C, Li C. Preparation and characterization of water-soluble chitosan produced by Maillard reaction. *Fish Sci.*, 2006; 72(5): 1096-1103.
- 148. Chen C-YU, Chung Y-C. Antibacterial effect of water-soluble chitosan on representative dental pathogens Streptococcus mutans and Lactobacilli brevis. *Journal of Applied Oral Science*. 2012; 20(6): 620-627.
- 149. Chung YC, Kuo CL, Chen CC. Preparation and important functional properties of water-soluble chitosan produced through Maillard reaction. *Bioresource Technol.*, 2005; 96(13): 1473-1482.
- 150. Crini G, Badot P-M. Application of chitosan, a natural aminopolysaccharide, for dye removal from aqueous solutions by adsorption processes using batch studies: A review of recent literature. *Progress in Polymer Science*, 2008; 33(4): 399-447.
- 151. Vakili M, Rafatullah M, Salamatinia B, Abdullah AZ, Ibrahim MH, Tan KB, Application of chitosan and its derivatives as adsorbents for dye removal from water and wastewater: A review. Carbohydrate polymers, 2014; 113(0): 115-130.
- 152. Ibrahim NA, Abou Elmaaty TM, Eid BM, Abd El-Aziz E. Combined antimicrobial finishing and pigment printing of cotton/ polyester blends. *Carbohydrate poly*mers, 2013; 95(1): 379-388.
- 153. Kalia S, Thakur K, Celli A, Kiechel MA, Schauer CL. Surface modification of plant fibers using environment friendly methods for their application in polymer composites, textile industry and antimicrobial activities: A review. *Journal of Environmental Chemical Engineering*, 2013; 1(3): 97-112.
- 154. Nayak R, Padhye R. Antimicrobial finishes for textiles. In: Paul R, editor. Functional Finishes for Textiles: Woodhead Publishing; 2015. p. 361-385.
- 155. Windler L, Height M, Nowack B. Comparative evaluation of antimicrobials for textile applications. *Environment international*. 2013; 53: 62-73.
- 156. Varona S, Martín Á, Cocero MJ. Formulation of a natural biocide based on lavandin essential oil by emulsification using modified starches. Chemical Engineering and Processing: Process Intensification, 2009; 48(6): 1121-1128.

- 157. Stupar M, Grbić ML, Džamić A, Unković N, Ristić M, Jelikić A. Antifungal activity of selected essential oils and biocide benzalkonium chloride against the fungi isolated from cultural heritage objects. South African Journal of Botany 2014; 93(0): 118-124.
- 158. Kim D, Jung S, Sohn J, Kim H, Lee S. Biocide application for controlling biofouling of SWRO membranes an overview. *Desalination*, 2009; 238(1–3): 43-52
- 159. Rajendran R, Radhai R, Kotresh TM, Csiszar E. Development of antimicrobial cotton fabrics using herb loaded nanoparticles. Carbohydrate polymers, 2013; 91(2): 613-617.
- 160. Nithya E, Radhai R, Rajendran R, Jayakumar S, Vaideki K. Enhancement of the antimicrobial property of cotton fabric using plasma and enzyme pre-treatments. *Carbohydrate polymers*, 2012; 88(3): 986-991.
- 161. Hebeish A, Hashem M, Shaker N, Ramadan M, El-Sadek B, Hady MA. Effect of post- and pre-crosslinking of cotton fabrics on the efficiency of biofinishing with cellulase enzyme. Carbohydrate polymers, 2009; 78(4): 953-960.
- 162. Nithya E, Radhai R, Rajendran R, Shalini S, Rajendran V, Jayakumar S. Synergetic effect of DC air plasma and cellulase enzyme treatment on the hydrophilicity of cotton fabric. Carbohydrate polymers, 2011; 83(4): 1652-1658.
- 163. Kim HW, Kim BR, Rhee YH. Imparting durable antimicrobial properties to cotton fabrics using alginate—quaternary ammonium complex nanoparticles. *Carbohydrate polymers*, 2010; 79(4): 1057-1062.
- 164. Hebeish A, El-Bisi MK, El-Shafei A. Green synthesis of silver nanoparticles and their application to cotton fabrics. International Journal of Biological *Macromolecules*, 2015; 72(0): 1384-1390.
- 165. Kittinaovarat S, Hengprapakron N, Janvikul W. Comparative multifunctional properties of partially carboxymethylated cotton gauze treated by the exhaustion or pad-dry-cure methods. Carbohydrate polymers, 2012; 87(1): 16-23.
- 166. Karapinar E, Phillips DAS, Taylor JA. Reactivity, chemical selectivity and exhaust dyeing properties of dyes possessing a 2-chloro-4-methylthio-striazinyl reactive group. *Dyes and Pigments*, 2007; 75(2): 491-497.
- 167. Kampyli V, Maudru E, Phillips DAS, Renfrew AHM, Rosenau T. Triazinyl reactive dyes for the exhaust dyeing of cotton: Influence of the oxido group on the reactivity of chloro and m-carboxypyridinium leaving groups. *Dyes* and Pigments. 2007; 74(1): 181-186.
- 168. Zhang L, Pornpattananangku D, Hu CM, Huang CM. Development of nanoparticles for antimicrobial drug delivery. Curr Med Chem., 2010; 17(6): 585-594.
- 169. Abdel-Mohsen AM, Abdel-Rahman RM, Hrdina R, Imramovský A, Burgert L, Aly

- AS. Antibacterial cotton fabrics treated with core—shell nanoparticles. *International Journal of Biological Macromolecules*, 2012; 50(5): 1245-1253.
- 170. Kayaoglu G, Ömürlü H, Akca G, Gürel M, Gençay Ö, Sorkun K, et al. Anti-bacterial Activity of Propolis versus Conventional Endodontic Disinfectants against Enterococcus faecalis in Infected Dentinal Tubules. *Journal of Endodontics*, 2011; 37(3): 376-381.
- 171. Patel J, Ketkar S, Patil S, Fearnley J, Mahadik K, Paradkar A. Potentiating antimicrobial efficacy of propolis through niosomal-based system for administration. *Integrative Medicine Research*, 2011; 13(0): 456-461.
- 172. Koo HR, P.L Cury, J.A Park, Y.K Bowen, W.H Effects of compounds found in propolis on Streptococcus mutans growth and on glucosyltransferase activity. *Antim Agents and Chemot.*, 2002; 46: 1302-1309.
- 173. Pinto MS, Faria JE, Message D, Cassini STA, Pereira CS, Gioso MM. Effect of green propolis extracts on patogenic bacteria isolated from milk of cows with mastitis. *Braz J Vet Res An Sci.*, 2004; 38: 278-283.
- 174. Loguercio AP, Groff ACM, Pedrozzo AF, Witt NM, Sae Silva AM, Vargras AC. In vitro activity of propolis extract against bovine mastitis bacterial agents. *Pesq. Agropec Bras.* 2006; 41(2): 347-349.
- 175. Sforcin JM, Fernandes Júnior A, Lopes CAM, Bankova V, Funari SRC. Seasonal effect on Brazilian propolis antibacterial activity. J of Ethnopharmacol., 2000; 73: 243-249.
- 176. Langoni H, Domingues PF, Funari SRC, Chande CG, Neves IR. In vitro antimicrobial effect of different endodontic materials and propolis on Enterococus faecalis. *Arq Bras Vet Zootec.*, 2006; 48: 227-229.
- 177. Valencia D, Alday E, Robles-Zepeda R, Garibay-Escobar A, Galvez-Ruiz JC, Salas-Reyes M. Seasonal effect on chemical composition and biological activities of Sonoran propolis. *Food Chemistry*, 2012; 131(2): 645-51.
- 178. Sharaf S, Higazy A, Hebeish A. Propolis induced antibacterial activity and other technical properties of cotton textiles. International Journal of Biological *Mac*romolecules, 2013; 59(0): 408-416.
- 179. Ashry El Sayed HEI, Ahmad TA. The use of propolis as vaccine's adjuvant. *Vaccine*, 2012; 31(1): 31-39.
- 180. Liu J, Willför S, Xu C. A review of bioactive plant polysaccharides: Biological activities, functionalization, and biomedical applications. *Bioactive Carbo*hydrates and Dietary Fibre, 2015; 5(1): 31-61.
- 181. Nelson G. Application of microencapsulation in textiles. *International Journal of Pharmaceutics*, 2003; 242: 66-72.
- 182. Freitas S, Merkle HP, Gander B. Microencapsulation by solvent extraction/ evaporation: reviewing the state of the

- art of microsphere preparation process technology. *Journal of controlled release*, 2005; 102(2): 313-332.
- 183. Sovilj V, Milanovic J, Katona J, Petrovic L. Preparation of microcapsules containing different contents of different kinds of oils by a segregative coacervation method and their characterization. Journal of the Serbian Chemical Society. 2010; 75(5): 615-627.
- 184. Sumithra M. Vasugi RN. Micro-encapsulation and nano-encapsulation of denim fabrics with herbal extracts. *Indian Journal of Fibre & Textile Research*, 2012; 37: 321-325.
- 185. Wang JM, Zheng W, Song QW, Zou H, Zhou Z. Preparation and characterization of natural fragrant microcapsules. *J Fibre Bioengin Informatics*. 2009; 1: 293-299.
- 186. Cheng SY, Yuan CWM, Kan CW, Cheuk KKL. Development of Cosmetic Textiles Using Microencapsulation Technology. *RJTA*, 2008; 12(4): 41-55
- 187. Achwal WB. Textiles with Cosmetics Substances. *Colourage*, 2003; 50(3): 21-42.
- 188. Mitali K, Maria NA. Emulsion-based techniques for encapsulation in biomedicine, food and personal care. Current Opinion in Pharmacology, 2014; 18: 47-55
- 189. Srinivasan K, Natarajan D, Mohanasundari C. Antibacterial, preliminary phytochemical and pharmacognostical screening on the leaves of Vicoaindica (L.)DC. J Pharmacol Ther. 2007; 6(1): 109–113.
- 190. Knez Ž, Markočič E, Leitgeb M, Primožič M, Knez Hrnčič M, Škerget M. Industrial applications of supercritical fluids: A review. *Energy*, 2014; 77(0): 235-243.
- 191. Yeo S-D, Kiran E. Formation of polymer particles with supercritical fluids: A review. The Journal of Supercritical Fluids, 2005; 34(3): 287-308.
- 192. Knez Ž, Škerget M, Knez Hrnčič M, Čuček D. Chapter 2 - Particle Formation Using Sub- and Supercritical Fluids. In: Fan VA, editor. Supercritical Fluid Technology for Energy and Environmental Applications. Boston: Elsevier; 2014. p. 31-67.
- 193. Türk M. Chapter 4 Formation of Organic Particles Using a Supercritical Fluid as Solvent. In: Michael T, editor. Supercritical Fluid Science and Technology. Volume 6: Elsevier; 2014. p. 57-75.
- 194. Finch CA, BR. Particle design using supercritical fluids: literature and patent survey. *Journal of Supercrit.*, 2012; 12: 156-162
- 195. Jennifer J, Michel P. Particle design using supercritical fluids- literature and patent survey, *Journal of Supercritical Fluids*, 2004; 20: 179-219.
- 196. Kumar AR, Rane YN. Encapsulation techniques. *International Dyer*, 2004; 189(7): 14-21.

- 197. Yan C, Zhang W. Chapter 12 Coacervation Processes. In: Gaonkar AG, Vasisht N, Khare AR, Sobel R, editors. Microencapsulation in the Food Industry. San Diego: Academic Press; 2014. p. 125-137.
- 198. Bilati U, Allémann E, Doelker E. Strategic approaches for overcoming peptide and protein instability within biodegradable nano- and microparticles. European Journal of Pharmaceutics and Biopharmaceutics, 2005; 59(3): 375-388.
- 199. Xie B. Preparation of uniform biodegradable microparticles using laser ablation. *International Journal of Pharma*ceutics, 2006; 325(1–2): 194-196.
- 200. Wen Y, Gallego MR, Nielsen LF, Jorgensen L, Everland H, Møller EH, Biodegradable nanocomposite microparticles as drug delivering injectable cell scaffolds. *Journal of Controlled Release*, 2011; 156(1): 11-20.
- 201. Ye M, Kim S, Park K. Issues in longterm protein delivery using biodegradable microparticles. *Journal of Controlled Release* 2010; 146(2): 241-60.
- 202. Krishnan S, Bhosale R, Singhal RS. Microencapsulation of cardamom oleoresin: Evaluation of blends of gum arabic, maltodextrin and a modified starch as wall materials. Carbohydrate polymers, 2005; 61(1): 95-102.
- 203. Butstraen C, Salaün F. Preparation of microcapsules by complex coacervation of gum Arabic and chitosan. Carbohydrate polymers. 2014; 99(0): 608-616.
- 204. Dima C, Cotârlet M, Alexe P, Dima S. Microencapsulation of essential oil of pimento [Pimenta dioica (L) Merr.] by chitosan/k-carrageenan complex coacervation method. *Innovative Food Science & Emerging Technologies*, 2014; 22(0): 203-211.
- 205. Beyki M, Zhaveh S, Khalili ST, Rahmani-Cherati T, Abollahi A, Bayat M. Encapsulation of Mentha piperita essential oils in chitosan–cinnamic acid nanogel with enhanced antimicrobial activity against Aspergillus flavus. *Industrial Crops and Products*, 2014; 54(0): 310-319.
- 206. Anitha A SRN, Joel D. Bumgardner SVN, and Rangasamy J. Approaches for Functional Modification or Cross□ Linking of Chitosan: Chitosan-Based Systems for Biopharmaceuticals. Delivery, Targeting and Polymer Therapeutics. 2012: 107-124.
- 207. Li M, Zhang G, Xu S, Zhao C, Han M, Zhang L. Cross-linked polyelectrolyte for direct methanol fuel cells applications based on a novel sulfonated cross-linker. *Journal of Power Sources*. 2014; 255(0): 101-117.
- 208. Wisnewski AV, Liu J, Redlich CA. Connecting glutathione with immune responses to occupational methylene diphenyl diisocyanate exposure. *Chemico-Biological Interactions*. 2013; 205(1): 38-45.

- 209. Shiotsuka RN. Hexamethylene Diisocyanate. In: Wexler P, editor. Encyclopedia of Toxicology (Third Edition). Oxford: Academic Press; 2014. p. 897-899
- 210. Li S, Jose J, Bouzidi L, Leao AL, Narine SS. Maximizing the utility of bio-based diisocyanate and chain extenders in crystalline segmented thermoplastic polyester urethanes: Effect of polymerization protocol. *Polymer*, 2014; 55(26) :6764-6775.
- 211. Sáfrány Á, Beiler B, László K, Svec F. Control of pore formation in macroporous polymers synthesized by single-step y-radiation-initiated polymerization and cross-linking. *Polymer*, 2005; 46(9): 2862-71
- 212. Harifi T, Montazer M. Past, present and future prospects of cotton cross-linking: New insight into nano particles. *Carbohydrate polymers*, 2012; 88(4): 1125-1140.
- 213. Biscarat J, Galea B, Sanchez J, Pochat-Bohatier C. Effect of chemical cross-linking on gelatin membrane solubility with a non-toxic and non-volatile agent: Terephthalaldehyde. *International Journal of Biological Macromolecules*, 2015; 74(0): 5-11.
- 214. Coimbra P, Gil MH, Figueiredo M. Tailoring the properties of gelatin films for drug delivery applications: Influence of the chemical cross-linking method. *International Journal of Biological Macromolecules*, 2014; 70(0): 10-19.
- 215. Teoh MM, Chung T-S, Wang KY, Guiver MD. Exploring Torlon/P84 co-polyam-ide-imide blended hollow fibers and their chemical cross-linking modifications for pervaporation dehydration of isopropanol. Separation and Purification Technology, 2008; 61(3): 404-413.
- 216. Wu W, Liu J, Cao S, Tan H, Li J, Xu F,. Drug release behaviors of a pH sensitive semi-interpenetrating polymer network hydrogel composed of poly(vinyl alcohol) and star poly[2-(dimethylamino)ethyl methacrylate]. International Journal of Pharmaceutics, 2011; 416(1): 104-109.
- 217. Matricardi P, Di Meo C, Coviello T, Hennink WE, Alhaique F. Interpenetrating Polymer Networks polysaccharide hydrogels for drug delivery and tissue engineering. Advanced Drug Delivery Reviews, 2013; 65(9): 1172-1187.
- 218. Kaity S, Isaac J, Ghosh A. Interpenetrating polymer network of locust bean gum-poly (vinyl alcohol) for controlled release drug delivery. *Carbohydrate polymers*, 2013; 94(1): 456-467.
- 219. Dragan ES. Design and applications of interpenetrating polymer network hydrogels. A review. Chemical Engineering Journal, 2014; 243(0): 572-590.
- 220. Syamili E, Elayarajah B, Kulanthaivelu, Rajendran R, Venkatrajah B, Ajith Kumar P. Antimicrobial cotton finish using green tea leaf extracts interacted with

- copper. *Asian J Textile*, 2012; 2(1): 6–12.
- 221. Straccia MC, Romano I, Oliva A, Santagata G, Laurienzo P. Crosslinker effects on functional properties of alginate/N-succinylchitosan based hydrogels. *Carbohydrate polymers*, 2014; 108(0): 321-330.
- 222. Russo R, Malinconico M, Santagata G. Effect of Cross-Linking with Calcium Ions on the Physical Properties of Alginate Films. *Biomacromolecules*. 2007; 8(10): 3193-3197.
- 223. DeFail AJ, Chu CR, Izzo N, Marra KG. Controlled release of bioactive TGF-β1 from microspheres embedded within biodegradable hydrogels. *Biomaterials*, 2006; 27(8): 1579-1585.
- 224. Jayaramudu T, Raghavendra GM, Varaprasad K, Sadiku R, Ramam K, Raju KM. lota-Carrageenan-based biodegradable Ag0 nanocomposite hydrogels for the inactivation of bacteria. Carbohydrate polymers, 2013; 95(1): 188-194.
- 225. Kevadiya BD, Rajkumar S, Bajaj HC, Chettiar SS, Gosai K, Brahmbhatt H, et al. Biodegradable gelatin–ciprofloxacin–montmorillonite composite hydrogels for controlled drug release and wound dressing application. *Colloids* and Surfaces B: Biointerfaces, 2014; 122(0): 175-183.
- 226. de Barros JC, da Conceicao ML, Neto NJ, da Costa AC, de Souza EL. Combination of Origanum vulgare L. essential oil and lactic acid to inhibit Staphylococcus aureus in meat broth and meat model. Brazilian journal of microbiology. Publication of the Brazilian Society for Microbiology, 2012; 43(3): 1120-1127.
- 227. Dimitrijevic SIM, Antonovic DG, Milanovic-Stevanovic MR, Mijin DZA. study of the synergistic antilisterial effects of a sub-lethal dose of lactic acid and essential oils from Thymus vulgaris L., Rosmarinusofficinalis L. and Origanumvulgare L. Food Chem Toxicol., 2007; 104: 774-782.
- 228. Sathianarayanan MP, Bhata NV, Kokate SS, Walunj VE. Antibacterial finish for cotton fabric from herbal products. *Indian Journal of Fibre & Textile Research*, 2010; 35: 50-58.
- 229. Banupriya J. Comparative Study on Antibacterial Finishes by Herbal and Conventional Methods on the Woven Fabrics. *Journal of Textile Science & Engineering*. 2013; 03(01): 123-128
- 230. Ghoranneviss M, Shahidi S, Anvari A, Motaghi Z, Wiener J, S'lamborova I. Influence of plasma sputtering treatment on natural dyeing and antibacterial activity of wool fabrics. *Prog Org Coat.*, 2010; 70(4): 388–893.
- 231. Chen C, Chang W. Antimicrobial activity of cotton fabric pretreated by microwave plasma and dyed with onion skin and onion pulp. *Indian Journal of Fibre & Textile Research*, 2007; 32: 122–125.

- 232. Kamel MM, El-Shishtawy R.M, Youssef B.M, Mashaly H. . Ultrasonic assisted dyeing. IV. Dyeing of cationised cotton with lac natural dye. *Dyes Pigm.*, 2007; 73(3): 279–284.
- 233. Vankar PS, Shanker R, Dixit S, Mahanta D, Tiwari SC. Sonicator dyeing of natural polymers with Symplocos spicata by metal chelation. *Fibers and Polymers*. 2008; 9(2): 121-127.
- 234. Ammayappan L, Moses JJ. Study of antimicrobial activity of aloevera, chitosan, and curcumin on cotton, wool, and rabbit hair. *Fibers Polym.*, 2009; 10(2): 161–166.
- 235. Nazarov VG, Stolyarov VP, Gagarin MV. Simulation of chemical modification of polymer surface. *Journal of Fluorine Chemistry*, 2014; 161(0): 120-127.
- 236. Muñoz-Bonilla A, León O, Cerrada ML, Rodríguez-Hernández J, Sánchez-Chaves M, Fernández-García M. Chemical modification of block copolymers based on 2-hydroxyethyl acrylate to obtain amphiphilic glycopolymers. European Polymer Journal. 2015; 62(0): 167-178.
- 237. Son Y, Ravikumar K, Kim B. . Development of berberine attraction sites onto cellulosic substrates modified by reactive bridging agent: statistical optimization and analysis. *Colloids Surf A.*, 2008; 325(3): 120–126.
- 238. Joshi M, Wazed Ali S, Rajendran S. Antibacterial finishing of polyester/cotton blend fabrics using neem (Azadirachta indica): a natural bioactive agent. *J Appl Polym Sci.* 2007; 106: 793–800.
- 239. Raja ASM, Thilagavathi G. Influence of enzyme and mordant treatments on the antimicrobial efficiency of natural dyes on wool materials. *Asian Journal of Textile*, 2011; 1(3): 138–144.
- 240. Kanth SV, Venba R, Jayakumar GC, Chandrababu NK. Kinetics of leather dyeing pretreated with enzymes: Role of acid protease. *Bioresource Technol.*, 2009; 100(8): 2430-2435.
- 241. Parvinzadeh M. Effect of proteolytic enzyme on dyeing of wool with madder. *Enzyme and Microbial Technology* 2007; 40(7): 1719-1722.
- 242. Khatri A, Peerzada MH, Mohsin M, White M. A review on developments in dyeing cotton fabrics with reactive dyes for reducing effluent pollution. *Journal* of Cleaner Production, 2015; 87(0): 50-57.
- 243. Jus S, Schroeder M, Guebitz GM, Heine E, Kokol V. The influence of enzymatic treatment on wool fibre properties using PEG-modified proteases. Enzyme and Microbial Technology. 2007; 40(7): 1705-1711.

- 244. Chakraborty JN. 31 Enzymatic dyeing of textiles. In: Chakraborty JN, editor. Fundamentals and Practices in Colouration of Textiles: Woodhead Publishing India; 2014. p. 418-432.
- 245. Kang S-M, Koh J, Noh S-Y, Kim S-J, Kwon Y-J. Effects of lugworm protease on the dyeing properties of protein fibers. *Journal of Industrial and Engineer*ing Chemistry, 2009; 15(4): 584-587.
- 246. Vigneswaran C, Ananthasubramanian M, Kandhavadivu P. 2 Industrial enzymes. In: Vigneswaran C, Ananthasubramanian M, Kandhavadivu P, editors. Bioprocessing of Textiles: Woodhead Publishing India; 2014. p. 23-52.
- 247. Mamun AA, Bledzki AK. Micro fibre reinforced PLA and PP composites: Enzyme modification, mechanical and thermal properties. Composites Science and Technology, 2013; 78(0): 10-17.
- 248. Arash I, Richard LG. Antimicrobial peptides. *Journal of the American Academy of Dermatology*, 2005; 52(3): 381-390.
- 249. Garry L, Sean PG, Brendan FG. The potential of antimicrobial peptides as biocides. Int. J. Mol. Sci. 2011; 12: 6566-6596.
- 250. Feng XX, Zhang LL, Chen JY, Zhang JC. New insights into solar UV-protective properties of natural dye. *Journal of Cleaner Production* 2007; 15(4): 366-372
- 251. Grifoni D, Bacci L, Zipoli G, Albanese L, Sabatini F. The role of natural dyes in the UV protection of fabrics made of vegetable fibres. *Dyes and Pigments* 2011; 91(3): 279-285.
- 252. Grifoni D, Bacci L, Di Lonardo S, Pinelli P, Scardigli A, Camilli F, et al. UV protective properties of cotton and flax fabrics dyed with multifunctional plant extracts. *Dyes and Pigments* 2014; 105(0): 89-96.
- 253. Kamel MM. El-Shishtawy RM, Yussef BM, Mashaly H. Ultrasonic assisted dyeing III. Dyeing of wool with lac as a natural dye. *Dyes Pigmnets* 2005; 65: 103–110.
- 254. Vankar PS, Shanker R, Srivastava J. Ultrasonic dyeing of cotton fabric with aqueous extract of Eclipta alba. *Dyes and Pigments* 2007; 72(1): 33–37.
- 255. Guesmi A, Ben Hamadi N, Ladhari N and Sakli F. Sonicator dyeing of modified acrylic fabrics with indicaxanthin natural dye. *Ind Crops Prod.* 2013; 42: 63–69.
- 256. Iqbal J, Bhatti IA and Adeel S. Effect of UV radiation on dyeing of cotton fabric with extract of henna leaves. *Indian Journal of Fibre & Textile Research* 2008; 33: 157–162.
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